

FORM PTO-1390
(REV 11-98)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

CHAIN=1B

U.S. APPLICATION NO. (If known, see 37 CFR 1.5)

09/402820

INTERNATIONAL APPLICATION NO.
PCT/US98/06900INTERNATIONAL FILING DATE
09 April 1998PRIORITY DATE CLAIMED
09 April 1997

TITLE OF INVENTION

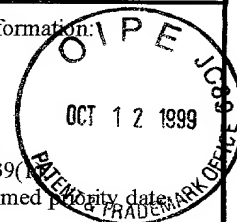
RECOMBINANT ANTIBODIES SPECIFIC FOR BETA-AMYLOID ENDS, DNA ENCODING AND...

APPLICANT(S) FOR DO/EO/US

Daniel G. CHAIN

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39.
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).



Items 11. to 16. below concern document(s) or information included:

11. ☒ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A FIRST preliminary amendment with substitute Sequence Listing and diskette.
☐ A SECOND or SUBSEQUENT preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information:
 1. A courtesy copy of the specification as originally filed.
 2. A small entity statement - small business concern.
 3. A courtesy copy of the first page of the International Publication (WO98/44955).
 4. A courtesy copy of the International Preliminary Examination Report.
 5. Formal drawings, 4 sheets, figures 1-5.

17. ☒ The following fees are submitted:**BASIC NATIONAL FEE** (37 CFR 1.492 (a) (1) - (5)) :

Neither international preliminary examination fee (37 CFR 1.482)
nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO
and International Search Report not prepared by the EPO or JPO \$970.00

International preliminary examination fee (37 CFR 1.482) not paid to
USPTO but International Search Report prepared by the EPO or JPO \$840.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but
international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$760.00

International preliminary examination fee paid to USPTO (37 CFR 1.482)
but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$670.00

International preliminary examination fee paid to USPTO (37 CFR 1.482)
and all claims satisfied provisions of PCT Article 33(1)-(4) \$96.00

ENTER APPROPRIATE BASIC FEE AMOUNT =

CALCULATIONS PTO USE ONLY

514 Rec'd PCT/PTO 12 OCT 1999

\$ 670.00

Surcharge of \$130.00 for furnishing the oath or declaration later than ☐ 20 ☐ 30
months from the earliest claimed priority date (37 CFR 1.492(e)).

\$

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total claims	22 - 20 =	2	X \$18.00	\$ 36.00	
Independent claims	3 - 3 =		X \$78.00	\$	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$260.00	\$	

TOTAL OF ABOVE CALCULATIONS =

\$ 706.00

Reduction of 1/2 for filing by small entity, if applicable. A Small Entity Statement
must also be filed (Note 37 CFR 1.9, 1.27, 1.28).

\$ - 353.00

SUBTOTAL =

\$ 353.00

Processing fee of \$130.00 for furnishing the English translation later than ☐ 20 ☐ 30
months from the earliest claimed priority date (37 CFR 1.492(f)).

\$

TOTAL NATIONAL FEE =

\$ 353.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be
accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +

\$

TOTAL FEES ENCLOSED =

\$ 353.00

Amount to be:

refunded

\$

charged

\$

a. ☒ A check in the amount of \$ 353.00 to cover the above fees is enclosed.

b. ☐ Please charge my Deposit Account No. _____ in the amount of \$ _____ to cover the above fees.
A duplicate copy of this sheet is enclosed.

c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any
overpayment to Deposit Account No. 02-4035. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

BROWDY AND NEIMARK, P.L.L.C.
419 Seventh Street N.W., Suite 300
Washington, D.C. 20004

SIGNATURE.

Roger L. Browdy

NAME

25,618

REGISTRATION NUMBER

Date of this submission: October 12, 1999

09/402820

514 Rec'd PCT/PTO 12 OCT 1999

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:)	Art Unit:
)	
Daniel G. CHAIN)	
)	Examiner:
IA No.: PCT/US98/06900)	
)	
IA Filed: April 9, 1998)	Washington, D.C.
)	
U.S. App. No.:)	
(Not Yet Received))	
)	October 12, 1999
Filing Date: October 12, 1999)	
)	
For: RECOMBINANT ANTIBODIES...)	Docket No.: CHAIN=1B

**PRELIMINARY AMENDMENT AND STATEMENTS IN SUPPORT OF FILING
AND SUBMISSIONS IN ACCORDANCE WITH 37 C.F.R. §1.821-1.825**

Honorable Commissioner of Patents and Trademarks
Washington, D.C. 20231

Sir:

Prior to the examination of the above-described application, please find attached a substitute Sequence Listing and please amend the present application as follows:

IN THE SPECIFICATION

Delete pages 45-47 in their entirety, and insert the attached new Sequence Listing, numbered pages 1-3, after the abstract.

IN THE CLAIMS

Please re-number pages 48-50 as pages 45-47.

09402820 101299

In re of CHAIN=1B

IN THE ABSTRACT

Please re-number the abstract as page 48.

REMARKS

Applicants have added into the present specification a substitute paper copy Sequence Listing section according to 37 C.F.R. §1.821(c) and §1.823(a). Furthermore, attached hereto is a 3 1/2" floppy disk containing the "Sequence Listing" in computer readable form in accordance with 37 C.F.R. §1.821(e).

The following statement is provided to meet the requirements of 37 C.F.R. 1.825(a) and 1.825(b).

I hereby state, in accordance with 37 C.F.R. §1.825(a), that the amendments included in the substitute sheets of the sequence listing are believed to be supported in the application as filed and that the substitute sheets of the sequence listing are not believed to include new matter.

I hereby further state, in accordance with 37 C.F.R. §1.825(b), that, on information and belief, the attached copy of the computer readable form is the same as the attached substitute paper copy of the sequence listing.

Applicants submit that the present application contains patentable subject matter and therefore urge the examiner to pass the case to issuance.

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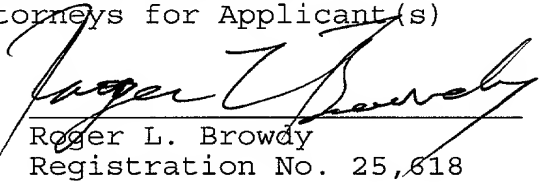
In re of CHAIN=1B

If the examiner has any questions or comments concerning the above described application, the examiner is urged to contact the undersigned at the phone number below.

Respectfully submitted,

BROWDY AND NEIMARK, P.L.L.C.
Attorneys for Applicant(s)

By


Roger L. Browdy
Registration No. 25,618

RLB:bcs

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09102820-10159
552707-02820460

Applicant or Patentee: Daniel CHAIN et al. Attorney's Docket No.: CHAIN-18
Appln. or Patent No.: _____ Filed or Issued: _____
For: RECOMBINANT ANTIBODIES SPECIFIC FOR BETA-AMYLOID ENDS, DNA ENCODING AND METHODS OF USE THEREOF

VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL BUSINESS ENTITY STATUS
(37 CFR 1.9(f) and 1.27(c)) - SMALL BUSINESS CONCERN

I hereby declare that I am

- ☐ I the owner of the small business concern identified below:
☐ I an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF SMALL BUSINESS CONCERN MINDSET LTD.
ADDRESS OF SMALL BUSINESS CONCERN Beit Eshel Street 1, 93227 Old Katamon, Israel

I hereby declare that the above identified small business concern qualifies as a small business concern as defined in 13 CFR 121.12, and reproduced in 37 CFR 1.9(d), for purposes of paying reduced fees to the United States Patent and Trademark Office, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention entitled RECOMBINANT ANTIBODIES SPECIFIC FOR BETA-AMYLOID ENDS, DNA ENCODING AND METHODS OF USE THEREOF by inventor Daniel G. CHAIN described in:

- ☒ the specification filed herewith with title listed as above.
☐ application no. _____, filed _____
☐ patent no. _____, issued _____

If the rights held by the above-identified small business concern are not exclusive, each individual, concern or organization having rights in the invention is listed below* and no rights to the invention are held by any person, other than the inventor, who would not qualify as an independent inventor under 37 CFR 1.9(c) if that person made the invention, or by any concern which would not qualify as a small business concern under 37 CFR 1.9(d), or a nonprofit organization under 37 CFR 1.9(e).

*Note: Separate verified statements are required from each named person, concern or organization having rights to the invention averring to their status as small entities (37 CFR 1.27).

NAME _____
ADDRESS _____

☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☐ NONPROFIT ORGANIZATION

NAME _____
ADDRESS _____

☐ INDIVIDUAL ☐ SMALL BUSINESS CONCERN ☐ NONPROFIT ORGANIZATION

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b))

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING Daniel Chain Dr DANIEL CHAIN
TITLE OF PERSON SIGNING OTHER THAN OWNER _____
ADDRESS OF PERSON SIGNING BEIT ESHEL SHAI 93227 OLD KATAMON ISRAEL
SIGNATURE Daniel Chain DATE 10-06-99

**RECOMBINANT ANTIBODIES SPECIFIC FOR BETA-AMYLOID ENDS, DNA ENCODING
AND METHODS OF USE THEREOF**

5

CROSS-REFERENCE TO RELATED APPLICATIONS

The present application claims priority under 35 USC
119(e) from U.S. provisional application no. 60/041,850,
10 filed April 9, 1997, the entire contents of which are hereby
incorporated by reference.

BACKGROUND OF THE INVENTION

Field of the Invention

15 The present invention relates to a method for
preventing or inhibiting progression of Alzheimer's Disease
through gene delivery to cells of the central nervous system.
The present invention also relates to a recombinant DNA
molecule containing a gene encoding a recombinant antibody
20 molecule end-specific for an amyloid- β peptide operably-
linked to a promoter capable of expressing a recombinant
antibody in cells of the central nervous system, and
pharmaceutical compositions thereof.

25 **Description of the Background Art**

A major histopathological hallmark of Alzheimer's
Disease (AD) is the presence of amyloid deposits within
neuritic and diffuse plaques in the parenchyma of the
amygdala, hippocampus and neocortex (Glenner and Wong, 1984;
30 Masters et al., 1985; Sisodia and Price, 1995). Amyloid is a
generic term that describes fibrillar aggregates that have a
common β -pleated structure. These aggregates exhibit
birefringent properties in the presence of Congo red and
polarized light (Glenner and Wong, 1984). The diffuse plaque
35 is thought to be relatively benign in contrast to the
neuritic plaque which appears to be strongly correlated with
reactive and degenerative processes (Dickson et al., 1988;
Tagliavini et al., 1988; Yamaguchi et al., 1989; Yamaguchi et
al., 1992). The principal component of neuritic plaques is a

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42 amino acid residue amyloid- β ($A\beta$) protein (Miller et al., 1993; Roher et al., 1993) that is derived from the much larger β -amyloid precursor protein, β APP (or APP) (Kang et al., 1987). $A\beta$ 1-42 is produced less abundantly than the 1-40 $A\beta$ peptide (Haass et al., 1992; Seubert et al., 1992), but the preferential deposition of $A\beta$ 1-42 results from the fact that this COOH-extended form is more insoluble than 1-40 $A\beta$ and is more prone to aggregate and form anti-parallel β -pleated sheets (Joachim et al., 1989; Halverson et al., 1990; Barrow et al., 1992; Burdick et al., 1992; Fabian et al., 1994). $A\beta$ 1-42 can seed the aggregation of $A\beta$ 1-40 (Jarrett and Lansbury 1993).

The APP gene was sequenced and found to be encoded on chromosome 21 (Kang et al., 1987). Expression of the APP gene generates several $A\beta$ -containing isoforms of 695, 751 and 770 amino acids, with the latter two β APP containing a domain that shares structural and functional homologies with Kunitz serine protease inhibitors (Kang et al., 1987; Kitaguchi et al., 1988; Ponte et al., 1988; Tanzi et al., 1988; Konig et al., 1992). The functions of β APP in the nervous system remain to be defined, although there is increasing evidence that β APP has a role in mediating adhesion and growth of neurons (Schubert et al., 1989; Saitoh et al., 1994; Saitoh and Roch, 1995) and possibly in a G protein-linked signal transduction pathway (Nishimoto et al., 1993). In cultured cells, β APPs mature through the constitutive secretory pathway (Weidemann et al., 1989; Haass et al., 1992; Sisodia 1992) and some cell-surface-bound β APPs are cleaved within the $A\beta$ domain by an enzyme, designated α -secretase, (Esch et al., 1990), an event that precludes $A\beta$ amyloidogenesis. Several studies have delineated two additional pathways of β APP processing that are both amyloidogenic: first an endosomal/lysosomal pathway generates a complex set of β APP-related membrane-bound fragments, some of which contain the entire $A\beta$ sequence (Haass et al., 1992; Golde et al., 1992); and second, by mechanisms that are not fully understood, $A\beta$ 1-40 is secreted into the conditioned medium and is present in cerebrospinal fluid in vivo (Haass et al., 1992; Seubert

et al., 1992; Shoji et al., 1992; Busciglio et al., 1993).
Lysosomal degradation is no longer thought to contribute
significantly to the production of A β (Sisodia and Price,
1995). The proteolytic enzymes responsible for the cleavages
5 at the NH₂ and COOH termini of A β termed β and γ ,
respectively, have not been identified. Until recently, it
was generally believed that A β is generated by aberrant
metabolism of the precursor. The presence, however, of A β in
conditioned medium of a wide variety of cells in culture and
10 in human cerebrospinal fluid indicate that A β is produced as
a normal function of cells.

If amyloid deposition is a rate-limiting factor to
produce AD, then all factors linked to the disease will
either promote amyloid deposition or enhance the pathology
15 that is provoked by amyloid. The likelihood of amyloid
deposition is enhanced by trisomy 21 (Down's syndrome)
(Neve et al., 1988; Rumble et al., 1989), where there is an
extra copy of the APP gene, by increased expression of APP,
and by familial Alzheimer's Disease (FAD)-linked mutations
20 (Van Broeckhoven et al., 1987; Chartier-Harlin et al., 1991;
Goate et al., 1989; Goate et al., 1991; Murrell et al., 1991;
Pericak-Vance et al., 1991; Schellenberg et al., 1992; Tanzi
et al., 1992; Hendricks et al., 1992; Mullan et al., 1992).
Some of these mutations are correlated with an increased
25 total production of A β (Cai et al., 1993; Citron et al.,
1992) or specific overproduction of the more fibrillogenic
peptides (Wisniewski et al., 1991; Clements et al., 1993;
Susuki et al., 1994) or increased expression of factors that
induce fibril formation (Ma et al., 1994; Wisniewski et al.,
30 1994). Collectively, these findings strongly favor the
hypothesis that amyloid deposition is a critical element in
the development of AD (Hardy 1992), but of course they do not
preclude the possibility that other age-related changes
associated with the disease, such as paired helical
35 filaments, may develop in parallel rather than as a result of
amyloid deposition and contribute to dementia independently.

The main focus of researchers and the principal aim
of those associated with drug development for AD is to reduce

the amount of A β deposits in the central nervous system (CNS). These activities fall into two general areas: factors affecting the production of A β , and factors affecting the formation of insoluble A β fibrils. A third therapeutic goal is to reduce inflammatory responses evoked by A β neurotoxicity.

With regards to the first, a major effort is underway to obtain a detailed understanding of how newly synthesized β APP is processed for insertion into the plasma membrane and to identify the putative amyloidogenic secretases that have been assigned on the basis of sites for cleavage in the mature protein. From a pharmacological perspective, the most direct way of reducing the production of A β is through direct inhibition of either β or γ secretase. No specific inhibitors are currently available although a number of compounds have been shown to indirectly inhibit the activities. Bafilomycin, for example, inhibits A β production with an EC₅₀ of about 50 nM (Knops et al., 1995; Haass et al., 1995), most likely through its action as an inhibitor of vacuolar H⁺ATPase co-localized in vesicles with the A β secretase. Another compound, MDL28170, used at high concentrations appears to block the activity of γ secretase (Higaki et al., 1995). It is generally hoped that the identification of the β or γ secretases might lead to the synthesis of specific protease inhibitors to block the formation of amyloidogenic peptides. It is not known, however, whether these enzymes are specific for β APP or whether they perhaps have other important secretory functions. Similarly, problems of target and targeting specificity will be encountered through any attempt to interfere with signal transduction pathways that may determine whether processing of β APP is directed through the amyloidogenic or non-amyloidogenic pathways. Moreover, these signal transduction mechanisms still need to be identified. In conclusion, present understanding of the complex and varied underlying molecular mechanisms leading to overproduction of A β offers little hope for selective targeting by pharmacological agents.

Given that neurotoxicity appears to be associated with β -pleated aggregates of A β , one therapeutic approach is to inhibit or retard A β aggregation. The advantage of this approach is that the intracellular mechanisms triggering the overproduction of A β or the effects induced intracellularly by A β need not be well understood. Various agents that bind to A β are capable of inhibiting A β neurotoxicity *in vitro*, for example, the A β -binding dye, Congo Red, completely inhibits A β -induced toxicity in cultured neurons (Yankner et al., 1995). Similarly, the antibiotic rifampacin also prevents A β aggregation and subsequent neurotoxicity (Tomiyama et al., 1994). Other compounds are under development as inhibitors of this process either by binding A β directly, e.g., hexadecyl-N-methylpiperidinium (HMP) bromide (Wood et al., 1996), or by preventing the interaction of A β with other molecules that contribute to the formation of A β deposition. An example of such a molecule is heparan sulfate or the heparan sulfate proteoglycan, perlecan, which has been identified in all amyloids and is implicated in the earliest stages of inflammation associated amyloid induction.

Heparan sulfate interacts with the A β peptide and imparts characteristic secondary and tertiary amyloid structural features. Recently, small molecule anionic sulfates have been shown to interfere with this reaction to prevent or arrest amyloidogenesis (Kisilevsky, 1995), although it is not evident whether these compounds will enter the CNS. A peptide based on the sequence of the perlecan-binding domain appears to inhibit the interaction between A β and perlecan, and the ability of A β -derived peptides to inhibit self-polymerization is being explored as a potential lead to developing therapeutic treatments for AD. The effectiveness of these compounds *in vitro*, however, is likely to be modest for a number of reasons, most notably the need for chronic penetration of the blood brain barrier.

As another means of inhibiting or retarding A β aggregation, WO 96/25435 discloses the potential for using a monoclonal antibody, which is end-specific for the free C-

terminus of the A β 1-42 peptide, but not for the A β 1-43 peptide, in preventing the aggregation of A β 1-42. While the administration of such an A β end-specific monoclonal antibody is further disclosed to interact with the free C-terminal residue of A β 1-42, thereby interfering with and disrupting aggregation that may be pathogenic in AD, there is no specific disclosure on how these A β C-terminal-specific monoclonal antibodies would be used therapeutically. Although direct or indirect manipulation of A β peptide aggregation appears to be an attractive therapeutic strategy, a possible disadvantage of this general approach may be that pharmacological compounds of this class will need to be administered over a long period of time, and may accumulate and become highly toxic in the brain tissue.

An alternative to a peptide-based approach is to elucidate the cellular mechanism of A β neurotoxicity and develop therapeutics aimed at those cellular targets. The focus has been on controlling calcium dysfunction of free radical mediated neuronal damage. It has been postulated that A β binds to RAGE (the receptor for advanced glycation end-products) on the cell surface, thereby triggering reactions that could generate cytotoxic oxidizing stimuli (Yan et al., 1996). Blocking access of A β to the cell surface binding site(s) might retard progression of neuronal damage in AD. To date there are no specific pharmacological agents for blocking A β -induced neurotoxicity.

In addition to therapeutic approaches through the direct administration of pharmacologically active agents, WO 89/01975 discloses a method of transplanting glial cells (actively secreting cells derived from within the brain), which have been transformed to express and secrete recombinant polymeric anti-acetylcholinesterase antibodies of the IgM class. It is predicted in the disclosure of WO 89/01975 that the antibody secreted by the transformed cells transplanted into the brain of a person suffering from Alzheimer's Disease may then alleviate or abolish the symptoms of the disease. This is a gene therapeutic approach arising from the observation that cells of the central

5 nervous system are very efficient in the secretion of
antibodies (Cattaneo and Neuberger, 1987). Piccioli et al.,
1991 and 1995, later demonstrated the ectopic neuronal
expression of recombinant antibodies from the promoter of the
neuronal *vgf* gene in a tissue-specific and developmentally
regulated manner. Thus, non-lymphoid cells, and in
particular, neuronal cells were found to be capable of
secreting functional immunoglobulins.

10 Citation of any document herein is not intended as
an admission that such document is pertinent prior art, or
considered material to the patentability of any claim of the
present application. Any statement as to content or a date
of any document is based on the information available to
applicant at the time of filing and does not constitute an
15 admission as to the correctness of such a statement.

SUMMARY OF THE INVENTION

20 The present invention relates to a novel method for
preventing the onset of Alzheimer's Disease or for inhibiting
progression of Alzheimer's Disease through the stable
expression in the brain of recombinant antibodies end-
specific for amyloid- β peptides. These ectopically expressed
recombinant antibody molecules, which are end-specific for
the N-terminus or C-terminus of amyloid- β peptides, prevent
25 the accumulation of amyloid- β peptides in the extracellular
space, interstitial fluid and cerebrospinal fluid and the
aggregation of such peptides into amyloid deposits in the
brain. Given the many possible mechanisms that might
contribute to the production of amyloid- β , coupled with the
30 tremendous diversity of interactions of $A\beta$ with the cell
surface and extracellular $A\beta$ -binding molecules capable of
bringing about chronic neurotoxicity, the present method is
directed to preventing the accumulation of $A\beta$ peptides in the
extracellular milieu of affected neurons as the focal point
35 of this heterogeneous pathological cascade. The present
invention also avoids the problems associated with the
repeated administration of pharmacological agents that
requires chronic penetration of the blood brain barrier.

It is therefore an object of the invention to overcome the deficiencies in the prior art by providing a novel method for preventing or inhibiting the progression of Alzheimer's Disease.

5 Another object of the invention is to provide a method whereby cells of the nervous system are conferred with the ability to ectopically express recombinant antibody molecules in the brain, which molecules are end-specific for the N-terminus or C-terminus of amyloid- β peptides, to
10 prevent the accumulation of amyloid- β peptides in the extracellular space, interstitial fluid and cerebrospinal fluid and the aggregation of such peptides into amyloid deposits in the brain.

A further object of the invention is to provide a
15 method for preventing or inhibiting the progression of Alzheimer's Disease by also inhibiting the interaction of amyloid- β peptides mediating amyloid- β induced neurotoxicity and inhibiting the amyloid- β induced complement activation and cytokine release involved in the inflammatory process
20 associated with Alzheimer's Disease.

Still another object of the invention is to provide a recombinant DNA molecule, containing a gene encoding a recombinant antibody molecule end-specific for the N-terminus or the C-terminus of an amyloid- β peptide and operably-linked
25 to a promoter which is expressed in the central nervous system.

Yet another object of the invention is to provide a vector for introducing the recombinant DNA molecule into cells of the central nervous system.

30 Still yet another object of the invention is to provide a pharmaceutical composition for preventing or inhibiting the progression of Alzheimer's Disease.

BRIEF DESCRIPTION OF THE DRAWINGS

35 Figure 1 shows a schematic representation of the β -amyloid precursor protein (β APP) and the products of α , β , and γ -secretase cleavage. The general locations of various domains are indicated along with the cleavage sites (α , β , γ)

for secretases. Figure 1 also schematically shows that the stable expression and secretion of ectopic A β -end-specific antibodies in the CNS inhibits (1) the accumulation of A β peptides and (2) the neurotoxic consequences of amyloid deposition without affecting the biological functions of the soluble β -amyloid precursor protein.

Figure 2 shows the amino acid sequence (SEQ ID NO:1) of the region in β APP from which β -amyloid peptides (A β) are derived. The arrows indicate the α -, β - or γ -secretase cleavage sites, and the amino acid residues corresponding to the synthetic peptides to be used as immunogens are indicated underneath the sequence by line segments.

Figures 3A-3D schematically show the structure of a whole antibody (Fig. 3A) with the variable domain of heavy (V_H) and light (V_L) chains and the constant domain(s) of light (C_L) and heavy (C_H1 , C_H2 , C_H3) chains, a Fab fragment (Fig. 3B), a Fv fragment (Fig. 3C), and a single chain Fv fragment (scFv) (Fig. 3D). The Fab fragment shown in Fig. 3B consists of a variable domain of heavy V_H and light V_L chain and the first constant domain (C_H1 and C_L) joined by a disulfide bridge. The Fv fragment shown in Fig. 3C represents the antigen binding portion of an antibody formed by a non-covalently linked variable region complex (V_H - V_L), whereas the single chain Fv shown in Fig. 3D joins the variable heavy V_H with the variable light V_L chain via a peptide linker.

Figure 4 schematically shows the construction of a scFv antibody by cloning the variable region of an end-specific anti-A β monoclonal antibody using the PCR amplification technique with primers A, B, C and D, and then joining together the variable heavy V_H chain and the variable light V_L chain with an interchain peptide linker (ICL). The shaded area represents hypervariable regions of the antigen binding site and LP designates the leader peptide of the heavy and light chains.

Figure 5 shows a schematic representation of the AAV ScFv α A β vector with the inverted terminal repeats (ITR),

human β APP promoter (Hu β APPP), SV40 polyadenylation signal (SV40pA) indicated. The plasmid backbone is pSSV9.

DETAILED DESCRIPTION OF THE INVENTION

5 The novel DNA molecules of the present invention contain a gene encoding a recombinant antibody molecule end-specific for the N-terminus or the C-terminus of an A β peptide. Such a recombinant antibody molecule discriminates between an A β peptide and the β -amyloid protein precursor
10 from which it is proteolytically derived, and is also referred to throughout the present specification as an "antisenilin". By "antisenilin" is meant a molecule which binds specifically to a terminus/end of an A β peptide to slow down or prevent the accumulation of amyloid- β peptides in the
15 extracellular space, interstitial fluid and cerebrospinal fluid and the aggregation into senile amyloid deposits or plaques and to block the interaction of A β peptides with other molecules that contribute to the neurotoxocity of A β .

 The method for preventing or inhibiting the
20 progression of Alzheimer's Disease in accordance with the present invention, involves delivering the gene encoding the antisenilin molecule into brain cells where antisenilins are then stably expressed and secreted into the extracellular space, interstitial fluid and cerebrospinal fluid. The
25 secretion of antisenilins into the extracellular space, interstitial fluid and cerebrospinal fluid, where soluble A β peptides are present, promotes the formation of soluble antisenilin-A β complexes. These soluble antisenilin-A β complexes are cleared from the central nervous system by
30 drainage of the extracellular space, interstitial fluid and cerebrospinal fluid into the general blood circulation through the arachnoid villi of the superior sagittal sinus. In this manner, soluble A β peptides are prevented from accumulating in the extracellular space, interstitial fluid
35 and cerebrospinal fluid to form amyloid deposits and/or to induce neurotoxicity (Fig. 1). Furthermore, clearance of soluble amyloid- β peptides in accordance with the present invention is expected to reduce the inflammatory process

observed in Alzheimer's Disease by inhibiting, for example, amyloid- β -induced complement activation and cytokine release, and block also the interaction of A β with cell surface receptors such as the RAGE receptor.

5 The composition of the present invention includes a recombinant DNA molecule containing an antisenilin gene in association with a means for gene delivery where this composition may be for use as a medicament for preventing or inhibiting the progression of Alzheimer's Disease.

10 As shown in Fig. 1 (see Schehr, 1994), and discussed in the Background Art section, the β -amyloid protein precursor (β APP) is believed also to serve as a precursor for a proteolytic product, soluble β -amyloid protein precursor (s β APP), thought to have growth promoting and neuroprotective functions. It will be readily
15 appreciated by those of skill in the art that the stable expression of antisenilins in the central nervous system will not interfere with the normal biological functions of β APP that are not associated with the formation of A β peptides.
20 In the novel recombinant DNA molecules of the present invention, the gene encoding an antisenilin molecule contains at least the nucleotide sequences which encode the antigen-binding domain of an end-specific monoclonal antibody molecule. Thus, the antisenilin molecule, which is a
25 recombinant antibody molecule containing the antigen-binding portion of a monoclonal antibody, is intended to encompass a chimeric or humanized immunoglobulin molecule of any isotype, as well as a single-chain antibody.

30 Chimeric antibodies are understood to be molecules, different portions of which are derived from different animal species, such as those having a variable region derived from a mouse monoclonal antibody and a human immunoglobulin constant region. Chimeric antibodies and methods for their production are well known in the art. For example, the DNA
35 encoding the variable region of the antibody can be inserted into or joined with DNA encoding other antibodies to produce chimeric antibodies (U.S. patent 4,816,567; Orlandi et al., 1989).

Single-chain antibodies as antisensilins can also be produced according to the present invention. These single chain antibodies can be single chain composite polypeptides having end-specific A β peptide binding capability and comprising a pair of amino acid sequences homologous or analogous to the variable regions of an immunoglobulin light and heavy chain (linked V_H-V_L or single chain Fv). Both V_H and V_L may copy natural monoclonal antibody sequences, or one or both of the chains may comprise a CDR-FR construct of the type described in U.S. Patent 5,091,513. The separate polypeptides analogous to the variable regions of the light and heavy chains are held together by a peptide linker. Methods of production of such single chain antibodies, e.g., single chain Fv (scFv), particularly where the DNA encoding the polypeptide structures of the V_H and V_L chains are characterized or can be readily ascertained by sequence analysis, may be accomplished in accordance with the methods described, for example, in U.S. Patent 4,946,778, U.S. Patent 5,091,513, U.S. Patent 5,096,815, Biocca et al., 1993, Duan et al., 1994, Mhashilkar et al., 1995, Marasco et al., 1993, and Richardson et al., 1995. Figures 3A-3D (from Biocca et al., 1995) schematically show an intact antibody (Fig. 3A), a Fab fragment (Fig. 3B), a Fv fragment consisting of a non-covalently linked variable region complex (V_H-V_L (Fig. 3C), and a single chain Fv antibody (Fig. 3D).

In constructing the recombinant gene encoding the antisensilin molecule, a hybridoma producing a monoclonal antibody end-specific for the N-terminus or C-terminus of an amyloid- β peptide is first obtained, where an end-specific antibody is defined as an antibody which uniquely recognizes the free N-terminus or the free C-terminus of a peptide and which can further discriminate between the peptide and the precursor from which it is proteolytically derived. The design of immunogenic peptides for use in immunization and the generation of monoclonal antibody producing hybridomas is based on similar peptides that have been previously used by several laboratories to generate monoclonal antibodies that uniquely recognize the free amino or carboxy-terminal of A β

(Harrington et al., 1993; Iwatsubo et al., 1994; Konig et al., 1996; Murphy et al., 1994; Gravina et al., 1995). While peptides of longer lengths have in some instances been used successfully to generate A β end-specific antibodies, Saido and co-workers (1993; 1994) established that there is a length of five amino acids for any given peptide which ensures that the specific free amino group at the N-terminus constitutes an essential part of the epitope recognized by the new antibody. Thus, a monoclonal antibody generated against an immunogenic peptide is evaluated for the selectivity of the antibody in its recognition of a free N- or C-terminus of an A β peptide. A competitive inhibition assay, using Enzyme-Linked Immunosorbant Assay (ELISA) or immunoprecipitation with peptides corresponding to different regions of A β and the region immediately preceding the β -secretase cleavage site in the extracellular domain of β APP, can determine the selectivity of the monoclonal antibody. When clearance of the amyloid peptides involved in the pathogenesis of Alzheimer's Disease, i.e., A β 1-40 (corresponding to residues 5-44 of SEQ ID NO:1), A β 1-42 (corresponding to residues 5-46 of SEQ ID NO:1), and A β 1-43 (corresponding to residues 5-47 of SEQ ID NO:1), is the major goal, then the monoclonal antibody is preferably end-specific for the N-terminus that is common to these A β peptides. In other cases, however, such as when used to treat a patient following the onset of Alzheimer's Disease, it may be preferable to select an antibody that will also interfere with the ability of A β peptides to seed aggregation or to interact with other molecules that either contribute to the seeding of A β deposition or mediate A β -induced cytotoxic effects. Immunogenic peptides of varying lengths, which incorporate either the free N-terminus or free C-terminus, are synthesized to allow for generating end-specific anti-A β antibodies and the recombinant DNA encoding for a recombinant A β end-specific antibody (antisenilin) used in a pharmaceutical composition for this type of selective application in preventing or inhibiting the progression of Alzheimer's Disease.

Those of skill in the art will appreciate that a cysteine residue can be added to the end of the above immunogenic peptides opposite from the end corresponding to the free N-terminus or the free C-terminus of A β peptides to facilitate coupling to a carrier protein. Keyhole limpet hemocyanin (KLH), ovalbumin and bovine serum albumin (BSA) are non-limiting examples of proteins that can be used as carriers for immunogens. The presence of an N-terminal or C-terminal cysteine residue on the synthetic immunogen peptides provides a free sulfhydryl group for covalent coupling to a maleimide-activated protein. A heterobifunctional reagent, such as an N-maleimido-6-aminocaproyl ester or a m-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS), is used to covalently couple the synthetic immunogenic peptide to the carrier protein (see for example, Hartlow, E. et al., Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. 1988). Commercial kits are also readily available for use in coupling peptide antigens to maleimide-activated large carrier proteins.

Monoclonal antibodies may be obtained by methods known to those skilled in the art. See, for example Kohler and Milstein, 1975; U.S. Patent No. 4,376,110; Ausubel et al., eds., Current Protocols in Molecular Biology, Greene Publishing Assoc. and Wiley Interscience, N.Y., (1987, 1992); and Harlow et al., *supra*; Colligan et al., eds., Current Protocols in Immunology, Greene Publishing Assoc. and Wiley Interscience, N.Y., (1992-1997), the contents of which references are incorporated entirely herein by reference.

Once monoclonal antibodies are generated, the selectivity and binding affinity (Kd) can be evaluated by ELISA, and *in vitro* bioassays can be performed on the antibodies to test for the efficacy of the A β end-specific monoclonal antibodies in blocking A β aggregation and A β -induced cytotoxicity as described below in Example 1. Preferably, these monoclonal antibodies have not only a selectivity that is end-specific for specific A β peptides, but also have a high binding affinity. It is intended that the DNA encoding any antibody that is end-specific for the N-

terminus or C-terminus of A β peptides and exhibits efficacy in blocking A β aggregation and A β induced cytotoxicity as described in Example 1 can be used in generating the recombinant antisenilin-encoding DNA molecules for use according to the present invention. For instance, the C-terminal end-specific monoclonal antibodies disclosed in WO 96/25435 may be used to obtain the recombinant antisenilin-encoding DNA molecules according to the present invention.

Messenger RNA (mRNA) may then be isolated from hybridomas producing A β end-specific monoclonal antibodies determined to be selective for the free N-terminus or free C-terminus of A β peptides. From the isolated hybridoma mRNA, cDNA is synthesized and the nucleotide sequence encoding the variable domains of the A β end-specific monoclonal antibody may then be cloned using the polymerase chain reaction (PCR) with primers based on the conserved sequences at each end of the nucleotide sequences encoding the V domains of immunoglobulin heavy chain (V_H) and light-chain (V_L). The presence of restrictions sites incorporated into the sequence of the PCR primers facilitates the cloning of PCR amplified products encoding the variable region of the appropriate chain.

A recombinant gene encoding a recombinant single chain Fv antibody molecule is constructed, for example, by joining nucleotide sequences encoding the V_H and V_L domains with a nucleotide sequence encoding a peptide interchain linker (Biocca et al., 1993; Duan et al., 1994; Mhashilkar et al., 1995; Marasco et al., 1993; Richardson et al., 1995; U.S. Patent 4,946,778; U.S. Patent 5,091,513, U.S. Patent 5,096,815) or by inserting the variable domain-encoding nucleotide sequences to replace the corresponding sequences encoding the variable domain in a human immunoglobulin gene to thereby encode for a recombinant chimeric antibody (Orlandi et al., 1989; U.S. Patent 4,816,567).

Standard reference works setting forth the general principles of recombinant DNA technology include Ausubel et al., eds., Current Protocols In Molecular Biology, Green Publishing Assoc. and Wiley Interscience, N.Y. (1987-1997),

Watson et al., Molecular Biology of the Gene, Volumes I and II, The Benjamin/Cummings Publishing Company, Inc., publisher, Menlo Park, CA (1987); Darnell et al., Molecular Cell Biology, Scientific American Books, Inc., publisher, New York, N.Y. (1986); Lewin, Genes II, John Wiley & Sons, publishers, New York, N.Y. (1985); Old et al., Principles of Gene Manipulation: An Introduction to Genetic Engineering, 2d edition, University of California Press, publisher, Berkeley, CA (1981); Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1989); and Berger et al., Guide to Molecular Cloning Techniques, Methods of Enzymology vo. 152, 1987, Academic Press, Inc. San Diego, CA. These references are hereby incorporated by reference.

The recombinant DNA molecule according to the present invention, which contains a recombinant antibody (antisenilin) gene, preferably also contains a promoter operably linked to the recombinant antisenilin gene and capable of expressing the antisenilin molecule in brain cells. It will also be appreciated that, in order to facilitate the secretion of the antisenilin molecule from transformed cells expressing antisenilin, a leader or signal peptide at the N-terminus is also provided.

A DNA molecule is said to be "capable of expressing" a polypeptide, such as the antisenilin molecule, if it contains nucleotide sequences which contain transcriptional and translational regulatory information, and such sequences are "operably linked" to nucleotide sequences which encode the polypeptide. An operable linkage is a linkage in which the regulatory DNA sequences and the DNA sequence sought to be expressed are connected in such a way as to permit gene expression. The regulatory regions needed for gene expression in general include a promoter region as well as the DNA sequences which, when transcribed into RNA, will signal the initiation of protein synthesis. Such regions will normally include those 5'-non-coding sequences involved with initiation of transcription and translation.

A promoter region would be operably linked to a DNA sequence if the promoter were capable of effecting transcription of that DNA sequence. As used herein, a "promoter sequence" is the sequence of the promoter which is found on the DNA and is transcribed by the RNA polymerase. Thus, to express antisenilins, transcriptional and translational signals recognized by the host cell are necessary.

The present method for preventing or inhibiting the progression of Alzheimer's Disease involves administering to a patient in need thereof a composition comprising a recombinant DNA molecule in association with means for gene delivery into cells of the central nervous system. The recombinant DNA molecule carries a gene encoding an antisenilin molecule operably-linked to a promoter where this operable linkage enables the expression of antisenilin molecules in the brain. The promoter is preferably a promoter which would follow the expression pattern of β APP with the highest level of expression in the hippocampus and cerebral cortex where amyloid deposition is most prevalent in Alzheimer's Disease. As a non-limiting example of a preferred promoter operably linked to the antisenilin gene, the thymidine kinase (Thy1) promoter has been shown to drive the expression of β APP in a region-specific manner that mimics the natural expression of β APP in the brain (Andra et al., 1996). Synapsin I promoter-based chimeric transgenes have been used to target expression of β APP in the CA subfields of the hippocampus and in the piriform cortex in brains of transgenic mice (Howland et al., 1995). A high level of β APP expression has been achieved in brain cortex of transgenic mice using a prion protein promoter (Hsiao et al., 1996). A number of advantages would be provided by using the β APP gene promoter to express the antisenilin gene. In particular, the antisenilin gene under the control of the β APP promoter would have identical anatomical and physiological patterns of expression as the β APP gene. The human β APP promoter has been characterized by a number of groups (e.g. Salbaum et al., 1988; La Fauci et al., 1989;

Wirak et al., 1991; Lahiri and Nall, 1995). The promoter has several regulatory domains including a heat-shock element and consensus sequences for the binding of transcription factors. Thus, expression of antisensilins under the control of the β APP gene can be enhanced as necessary in specific regions of the brain by applying any of a number of inducing agents, for example, growth factors, retinoic acid, and interleukin-1. A preproenkephalin promoter has also been reported to yield region-specific and long term expression in an adult rat brain after direct *in vivo* gene transfer (Kaplitt et al., 1994).

In order to facilitate the introduction of a recombinant DNA molecule carrying an antisensilin gene operably-linked to a promoter into cells of the central nervous system, a number of different means for gene delivery can be used in association with the recombinant DNA molecule. The term "means for gene delivery" is meant to include any technique suitable for delivery of DNA molecules across the blood brain barrier and/or for transmembrane delivery across cell membranes. Non-limiting examples of the means for gene delivery are viral vectors (e.g., adeno-associated virus-based vectors), lipids/liposomes, ligands for cell surface receptors, etc.

The recombinant DNA molecule carrying the antisensilin gene is associated with the means for gene delivery where such association is intended to encompass, for example, the situation in which the means for gene delivery in a viral vector and the antisensilin gene is incorporated in the DNA of the viral vector or packaged in the viral particle; the situation in which the means for gene delivery is a liposome and the antisensilin gene is complexed therewith; the situation in which the means for gene delivery is a ligand for a cell surface receptor and the antisensilin gene is conjugated or otherwise bound thereto; etc. Thus, "in association with" includes incorporating or packaging in, complexing with, conjugating or binding to, and any other manner of associating the antisensilin gene with the means for gene delivery. It will be appreciated that the recombinant

DNA molecule may be in association with more than one means for gene delivery, particularly where the recombinant DNA molecule is to be delivered across both the blood brain barrier and the cell membrane of brain cells.

5 Adeno-associated virus (AAV) was initially isolated as a tissue culture contaminant and was later found as a non-pathogenic coinfecting agent during an adenovirus outbreak in children (Blacklow et al., 1968). It is a single-stranded DNA virus of the parvovirus group with a 4.7 kb genome. As
10 one of the smallest human DNA viruses, AAV requires coinfection with a helper virus, usually an adenovirus or herpesvirus, for efficient replication in order to complete its life cycle (Carter, 1990). In the absence of helper virus infection, AAV becomes latent and stably integrates at
15 high frequency, often at a specific site on chromosome 19 (Kotin et al., 1990; 1991; 1992; Samulski et al., 1991). The AAV genome has been sequenced and it was discovered that the sole sequence needed for integration of an AAV vector is in the terminal 145 nucleotide inverted terminal repeats (ITR),
20 thus making the cloning capacity nearly 4.7 kb (Muzyczka, 1992). Due to the non-pathogenic nature of the virus, its broad host cell range, and its ability to take advantage of a natural mechanism for high frequency integration, AAV is particularly suitable as a vector for gene delivery/transfer
25 into cells. Moreover, while conventional retroviruses have a requirement for genomic DNA synthesis, AAV vectors have a unique ability to introduce foreign genes into non-dividing or quiescent cells. These characteristics are being increasingly exploited for gene expression in the mammalian
30 brain, and several genes related to Alzheimer's Disease have been expressed in the brain using AAV vectors (Makimura et al., 1996). Recent studies by Du et al., 1996, indicate that AAV vectors can efficiently transduce and stably express a foreign gene, e.g., *lacZ*, in post-mitotic human neurons. The
35 expression of foreign genes in neuronal cells has also been reported using liposome-mediated transfection with AAV-derived plasmids (Meyer et al., 1995; Wu et al., 1994, 1995).

Low et al., U.S. Patent 5,108,921, reviews available methods for transmembrane delivery of molecules such as proteins and nucleic acids by the mechanism of receptor mediated endocytotic activity. These receptor systems include those recognizing galactose, mannose, mannose-6-phosphate, transferrin, asialoglycoprotein, transcobalamin (vitamin B₁₂), α -2 macroglobulins, insulin and other peptide growth factors such epidermal growth factor (EGF). Low et al. also teaches that nutrient receptors, such as receptors for biotin and folate, can be advantageously used to enhance transport across the cell membrane due to the location and multiplicity of biotin and folate receptors on the membrane surfaces of most cells, and the associated receptor mediated transmembrane transport processes. Thus, a complex formed between a compound to be delivered into the cytoplasm and a ligand, such as biotin or folate, is contacted with a cell membrane bearing biotin or folate receptors to initiate the receptor mediated trans-membrane transport mechanism and thereby permit entry of the desired compound into the cell.

A biotin ligand can be attached to a DNA molecule, for example, by incorporating commercially available biotinylated deoxynucleotide triphosphates, e.g., biotin-14-dATP or biotin-14-dCTP from Life Technologies, Inc., Gaithersburg, MD, using terminal deoxynucleotidyl transferase (Karger, B.D., 1989). Biotin-14-dATP is a dATP analog with biotin attached at the 6-position of the purine base by a 14-atom linker and biotin-14-dCTP is a dCTP analog with biotin attached at the N⁴-position of the pyrimidine base also by a 14-atom linker.

Whether incorporated into a viral-based or plasmid vector for packaging into a virus, attached to a neural receptor-binding ligand molecule, complexed with cationic lipids or cationic liposomes, or in association with other suitable means for gene delivery, the recombinant DNA molecule encoding an antisense gene operably linked to a promoter is administered to a subject by injection. Stereotactic microinjection into different brain regions

through use of established coordinates can be used to deliver the viral packaged or ligand-bound recombinant DNA molecule directly into the extracellular environment, e.g., cerebrospinal fluid, surrounding brain cells for subsequent transmembrane delivery into the cells themselves.

As direct injection into the brain is an invasive procedure, it is preferred that the viral packaged or ligand-bound recombinant DNA molecule be administered by intravenous or intra-arterial injection. The viral packaged or ligand-bound recombinant DNA can further be in association with other means for gene delivery, such as to effect gene delivery across the blood-brain barrier into the central nervous system. Zhu et al., 1993, demonstrated that cationic lipid-plasmid DNA complexes can be delivered systemically to all tissues including the brain. Recently, it has also been shown that intra-arterially administered cationic liposomes containing the thymidine kinase gene was successful in a rat model of brain tumor where regression was achieved without apparent toxicity or histological damage (Laine et al., 1995). Gene delivery by liposomes is well covered in the scientific literature and in patent publications, and extensively reviewed by Lasic, D.D., In: Liposomes in Gene Delivery, CRC Press, Boca Raton, Florida, 1997, which is hereby incorporated entirely by reference.

Once delivered to the brain, the viral packaged recombinant DNA molecule, either ligand-bound or in association with another suitable means for gene delivery, transforms brain cells, which subsequently express antisenilin molecules (recombinant antibody molecules end-specific for A β peptides) and secrete the expressed antisenilins into the extracellular space, interstitial fluid and cerebrospinal fluid. The secreted antisenilins then form a soluble complex with A β peptide to which they are end-specific in the extracellular space, interstitial fluid and cerebrospinal fluid. These soluble antisenilin-A β peptide complexes prevent the aggregation of A β peptides into amyloid deposits and prevent A β -induced neurotoxicity by clearing A β peptides from the central nervous system through drainage of

the extracellular space, interstitial fluid and cerebrospinal fluid into the general blood circulation where they will be eliminated by protease digestion. Accordingly, the accumulation of newly-secreted soluble A β peptides responsible for amyloid deposition and A β -induced neurotoxicity is prevented.

While the present method for preventing or inhibiting the progression of Alzheimer's Disease is intended to be primarily used for patients with a clear genetic disposition to developing Alzheimer's Disease, it can also be used prophylactically to "immunize" the population in general against the occurrence of such a prevalent and debilitating disease. The preferred route of administration is intravenous or intra-arterial. However, despite the invasiveness of microinjection directly into regions of the brain, this route of administration is intended to be within the scope of the invention. In particular, patients having Down's Syndrome or familial Alzheimer's Disease-linked mutations who are expected to develop Alzheimer's Disease due to their predisposition or patients who already suffer from Alzheimer's Disease can be treated by direct microinjection into the brain. The benefit of this treatment is expected to outweigh the risks of an invasive technique such as injection into the brain.

The recombinant DNA molecule which contains an antisenilin gene in association with a means for gene delivery may be used in the preparation or manufacture of a medicament/pharmaceutical composition. The pharmaceutical compositions contain an amount of the recombinant DNA molecule effective to achieve its intended purpose. For instance, when the means for gene delivery is a viral vector, such as an AAV vector, a suitable dosage of viral particles in a pharmaceutical composition to be stereotactically microinjected into different locations in the brain is in the range of about 5×10^4 to 1×10^{11} particles. When a ligand, such as biotin, is used as the means for gene delivery by administration directly into the brain, ligand-bound DNA molecules in the range of about 0.5 to 100 μ g are suitably

used. For such ligand bound DNA molecules, it is preferred that the DNA molecules are condensed beforehand to protect these molecules in the extracellular milieu of cells within the central nervous system. Pharmaceutical compositions and dosages of DNA molecules complexed with cationic lipids or cationic liposomes are discussed in Lasic, 1997, *supra*. Furthermore, the pharmaceutical compositions may contain suitable pharmaceutically acceptable excipients, such as are well-known in the art.

Having now generally described the invention, the same will be more readily understood through reference to the following prophetic example, which is provided by way of illustration and is not intended to be limiting of the present invention.

EXAMPLE 1

The strategy and the protocols for use in developing recombinant DNA molecules containing a gene encoding a recombinant antisenilin antibody molecule end-specific for an amyloid- β peptide are described below.

MONOCLONAL A β END-SPECIFIC ANTIBODY PRODUCTION

Immunogen peptide synthesis

Several peptides of varying lengths incorporating either the free N-terminus or free C-terminus are prepared using an Applied Biosystems Peptide Synthesizer (430A). The synthetic peptides are purified by HPLC and characterized using both amino acid composition and NH₂-terminal micro sequence analyses.

Peptide N1/5 A β _{1-40/42} (mAb: "Antisenilin N1/5")

A peptide corresponding to the first five amino acid residues of A β (1-40 and 1-42), as schematically represented by the appropriate line segment in Fig. 2, is synthesized. The peptide contains a cysteine residue at the C terminus and has the sequence of SEQ ID NO:2 (See Figure 1).

Peptide N1/7A β _{1-40/42} (mAb: "Antisenilin N1/7")

A peptide corresponding to the first seven amino acid residues of A β (1-40 and 1-42), as schematically represented by the appropriate line segment in Fig. 2, are synthesized. The peptide contains a cysteine residue at the C terminus and has the sequence of SEQ ID NO:3.

Peptide C34/40A β ₁₋₄₀ (mAb: "Antisenilin C34/40")

A peptide corresponding to the last seven amino acid residues of A β (1-40), as schematically represented by the appropriate line segment in Fig. 2, is synthesized. The peptide contains a cysteine residue at the N-terminus and has the sequence of SEQ ID NO:4.

Peptide C36/42A β ₁₋₄₂ (MAb: "Antisenilin C36/42")

A peptide corresponding to the last seven amino acid residues of A β (1-42), as schematically represented by the appropriate line segment in Fig. 2, is synthesized. The peptide contains a cysteine residue at the N-terminus and has the sequence of SEQ ID NO:5.

Peptide conjugation

The purified peptides are conjugated to bovine serum albumin (BSA) using N-maleimido-6-aminocaproyl ester of 1-hydroxyl-2-nitro-4-benzene-sulfonic acid.

Immunization and hybridoma monoclonal antibody production

Phase 1: Four sets of 10 Balb/c mice are immunized with the purified BSA-conjugated peptides described above using standard immunization protocols (Taggart and Samloff, 1983).

Phase 2: Following the completion of the immunization protocol, a fusion procedure is performed using spleenocytes from the hyperimmunized mice and an appropriate myeloma cell-line SP2/0-Ag14 (ATCC CRL 1581), NS-1 (ATCC TIB18), or equivalent. This procedure is performed using polyethylene glycol, and the selection of successful fusion products are achieved by means of HAT media. Viable hybridoma colonies are grown out in 96 well plates.

Phase 3: Screening of all wells containing successful fusion products are carried out using ELISA described in the next section with the peptide antigens. Supernatants from several wells are also screened in the in vitro bioassays as described below.

Phase 4: On the basis of the results of ELISA assays and the evaluations based the results of the bioassays, subcloning is performed by limiting dilutions on the selected colonies.

10 ELISA detection and affinity determinations

The specificity and binding affinities (Kds) of the monoclonal antibodies are evaluated by ELISA assays (Engvall and Perlmann, 1971) using a set of synthetic peptides corresponding to A β 1-42, A β 1-40, and residues 1-52, 1-11, -2[KM]-11, -1[M]-11, 1-28, 35-40, 35-42, and 35-44 found in A β peptides and β APP from which they are derived. In addition, the immunogenic peptide sequences, corresponding to the N-terminus or C-terminus of A β peptides, and conjugated to a different carrier protein, such as keyhole limpet hemocyanin (KLH) and ovalbumin, are used to determine whether the resultant monoclonal antibodies are end-specific for A β peptides and non-specific for the carrier protein or the cysteine bridge.

To test the protocol to be used subsequently to generate monoclonal antibodies, high affinity polyclonal antibodies specific for the free N-terminus of A β peptides were made where the antibodies were raised using the restricted peptide: H₂N - SEQ ID NO:6 - aminohexanoate-C-amide. The peptides were synthesized using solid phase Fmoc chemistry. The peptides were then cleaved and analyzed by mass spectroscopy and high performance liquid chromatography (HPLC). HPLC purification was achieved using a C-18 YMC column (10 μ packing, 120 A pore size, 10 X 250 mm) in a buffer system of A: H₂O/0.1% TFA and B:CH₃CN/0.08% TFA. The appropriate fractions were pooled, lyophilized, and again subjected to mass spectroscopy and HPLC analysis. The peptide was coupled to KLH for immunization, BSA for ELISA detection, with the cross-linker MBS. Rabbits were immunized

at 3 week intervals, and the titer assessed by ELISA using acetal-SEQ ID NO:7-Ahx-C-amide. This peptide corresponds to a sequence of amino acid residues that spans the 0 to 1 splice site that yields the free N-terminus of A β peptides.

5 The same spanning peptide was coupled to a thiol coupling gel via their cysteine residue and used to preabsorb away all antibodies which do not depend upon the free amine-Asp being present. The antibodies were then purified and collected using the N-terminal peptide. Whereas the crude serum shows

10 substantial activity towards the spanning peptide, once affinity purified, there is no reactivity of the resulting antibody with the spanning peptide, only with the N-terminal peptide.

To generate monoclonal antibodies specific for the

15 N-terminus of the A β peptides, mice are immunized at 3 week intervals using: H₂N-SEQ ID NO:6-aminohexanoate-C-amide conjugated to BSA prepared as described for the preparation of polyclonal. The titer in each mouse is also assessed by ELISA as described above. After spleen cell fusion of the

20 mice containing the highest titer, several clones are isolated and screened using the spanning peptide ELISA detection method.

25 In vitro bioassays to test efficacy of A β end-specific antibodies in blocking A β aggregation and A β -induced cytotoxicity

A) Effects on A β Fibril Formation: As shown by Jarrett et al. (1993), the carboxyterminus of A β is critical for the

30 "seeding" of amyloid formation which is probably responsible for the greatly accelerated rate of amyloid plaque formation in Alzheimer's Disease (Yankner and Mesulam, 1991). Amyloid formation by the kinetically soluble peptides, such as A β 1-40, can be nucleated or "seeded" by peptides such as A β 1-42

35 that include the critical C-terminal residues 41(Ile) and 42(Ala). After the April 9, 1997, filing date of the provisional U.S. application on which the present application claims benefit of priority, abstracts by Solomon et al.

5 (1997) and Frenkel et al. (1997) reported that their studies show that antibodies directed towards the N-terminal region of positions 1-16 of A β peptides bind to formed fibrils and lead to disaggregation. The anti-aggregating epitope of such an antibody is reported to be located within just the four amino acids Glu Phe Arg His (SEQ ID NO:8) of residue positions 3-6. These four amino acid residues of SEQ ID NO:8 are all present in the immunizing peptides for the N-terminus of A β . The ability of C-terminus or N-terminus end-specific A β antibodies to block seeding by A β 1-42 or to prevent aggregation of amyloid peptides is tested using standard aggregation assays (Wood et al., 1996). The A β 1-40 peptide is solubilized to 5 mg/ml in 1,1,1,3,3,3-hexafluoro-2-propanol. The peptide is concentrated to dryness and resolubilized in phosphate-buffered saline (PBS), pH 7.4, to a final concentration of 230 μ M. A solution of A β 1-42 (20 μ M) is stirred for 3 days and sonicated for 30 min to produce amyloid fibrils. Preaggregated A β 1-42 at 2nM concentration is added to the supersaturated pH 7.4 incubation to seed aggregation of A β 1-40. Aggregate formation in the absence and in the presence of each A β end-specific monoclonal antibody is determined by monitoring the turbidity of samples prepared in microtiter wells using a microtiter plate reader at 405 nm. The reaction is also monitored by thioflavin-T fluorescence as described by Wood et al. (1996). The ability of N-terminus-specific antibodies to promote disaggregation of amyloid peptide fibrils is tested by testing the displacement of [125 I]-labeled amyloid aggregated peptides from a collagen matrix containing non-aggregated peptides coated onto 96-well microtiter-plastic-coated plates. In addition, the ability of N-terminus-specific-antibodies to protect neurons from A β -induced damage is assessed by the trypan blue exclusion method, intracellular calcium measurements, scanning and transmission electron microscopy and by confocal microscopy.

B) A β -induced neurotoxicity: The receptor for advanced glycation end products (RAGE) mediates some of the neurotoxic

effects of A β on neurons and microglia (Yan et al., 1996). End-specific antibodies are tested for their ability to inhibit the receptor-mediated neurotoxicity by competitive inhibition. The antibodies are tested both with purified
5 RAGE receptor preparations and by measuring their effect on A β -induced cellular oxidant stress.

The RAGE receptor is purified from a bovine lung extract dissolved in tris-buffered saline containing octyl- β -glucoside (1%) and phenylmethylsulfonylfluoride (2 mM) and
10 applied to a heparin hyperD column (Biosepra). The column is eluted with a gradient of NaCl and fractions with maximal binding of ¹²⁵I-labeled A β are identified. The fractions are pooled and loaded onto hydroxyapatite ultragel (Biosepra) and eluted with increasing concentrations of phosphate.

Fractions with maximal binding of ¹²⁵I-labeled A β are applied
15 to preparative non-reduced polyacrylamide SDS gels (10%). The RAGE receptor protein M_r 50,000 is identified by Coomassie Blue staining and the region in adjacent lanes are cut and eluted. Competitive inhibition by the end-specific
20 antibodies to binding of ¹²⁵I-labeled A β (1-40/1-42) to the RAGE receptor is determined in a number of ways: (1) different amounts (0-150 μ g) of purified protein are immobilized on microtiter wells and incubated with 100 nM
25 ¹²⁵I-labeled A β (1-40/1-42); (2) different amounts (0-250 nM) of ¹²⁵I-labeled A β (1-40/1-42) are incubated in microtiter wells pre-coated with 5 μ g purified RAGE receptor; and (3) different amounts (0-500 μ g/ml) of A β (1-40/1-42) are
30 immobilized on microtiter wells and incubated with 50 nM ¹²⁵I-labeled RAGE receptor. In each assay, the amount of ligand binding to the well in the presence of different amounts of antibody is determined by counting the amount of radioactivity in the wells with a gamma-scintillation counter.

To evaluate the efficacy of the different
35 end-specific A β monoclonal antibodies as inhibitors of A β -induced cellular oxidant stress, cultured mouse brain microvascular endothelial cells (Breitner et al., 1994) are incubated with 0.25 μ M A β in the presence of different

amounts of the antibodies, and cellular oxidant stress is assessed by measuring the dose-dependent generation of thiobarbituric acid-reactive substances using the TBARS assay as previously described (Dennery et al., 1990; Yan et al., 1996). In a parallel assay system (developed by Khoury et al., 1996), the inhibitory effects of the antibodies are tested on A β -induced production of oxygen-reactive species in N9 mouse microglial cells. N9 cells (5×10^4) are incubated at 37°C in the presence of different amounts of the antibodies in 50 μ l PD-BSA (phosphate-buffered saline lacking divalent cation having 1 mg/ml BSA) containing 1 μ M H₂DCF (2',7'-dichlorofluorescein diacetate), a dye that fluoresces upon oxidation (Wan et al., 1993) on multispot slides coated with A β peptides. At various time points, aliquots of the culture medium are taken and the fluorescence is measured in a fluorescence plate reader (Cytofluor II).

C) Effect on interactions with proteoglycans: The vascular cell derived heparan sulfate proteoglycan, perlecan, has been identified in all amyloid deposits and is implicated in the earliest stages of inflammation-associated amyloid induction through high-affinity binding interactions with A β (Snow et al. 1989; 1995). Binding of perlecan to A β imparts secondary and tertiary amyloid structural features which suggest that molecules that interfere with the interaction may prevent or arrest amyloidogenesis.

End-specific A β monoclonal antibodies made to peptides of different lengths that correspond to the N-terminus of the peptide are evaluated for their ability to block the binding of perlecan to the perlecan binding site in the N-terminus region of A β (Snow et al., 1995). These evaluations are based on a solid-phase binding assay using perlecan isolated from cultured endothelial cells prepared from calf thoracic aortas as described in detail by (Snow et al. 1995). Polyvinyl micro-titer wells are coated with 100 μ l of nitrocellulose solution and allowed to dry. Wells are then coated overnight at room temp with unlabeled perlecan to give 0.28 μ g of bound perlecan per well, and blocked

overnight at room temp with 200 μ l of 5% non-fat dried milk. Various quantities of 125 I A β (7000 cpm/pM) diluted in 100 μ l of TBS/0.05% Tween 20 (TBST) are added in triplicate to the wells and incubated for 2.5 h at room temp on an orbital shaker. At the end of the incubation period, free 125 I A β is removed with six washes of TBST. Bound 125 I is extracted in 100 μ l 1N sodium hydroxide and "bound" versus "free" radioactivity is quantitated by liquid scintillation counting. Scatchard analysis is performed after incubating 125 I-A β in the presence of increasing amounts of monoclonal antibody.

CLONING AND ASSEMBLY OF RECOMBINANT GENES

mRNA isolation and cDNA synthesis from hybridomas

Messenger RNA (mRNA) is prepared from 5×10^8 hybridoma cells as described by Griffiths and Milstein (1985). First-strand cDNA synthesis is performed according to standard procedures (Maniatis et al., 1989).

PCR amplification, cloning of variable antigen-binding region and construction of single-chain antibodies

Techniques have been developed for the cloning of immunoglobulin variable domains from genomic DNA and cDNA using the polymerase chain reaction (Orlandi et al., 1989; Ward et al., 1989; Richardson et al., 1995). Primers based on conserved sequences at each end of the nucleotide sequences encoding V domains of mouse immunoglobulin heavy-chain (V_H) and kappa light-chain (V_K) also incorporate restriction sites that permit force-cloning of the amplified product containing the variable region of each chain. These primers are capable of amplifying most immunoglobulin mRNA of the mouse repertoire.

As shown in Fig. 4, PCR on cDNA from the hybridoma cells is performed using the primers described by Richardson et al., 1995. The scFv antisensilin gene is assembled from the amplified DNA corresponding to the V_H and V_L regions and an interchain linker, expressed in *E. coli*, and reamplified

by PCR using primers that incorporates a stop codon at the 3'-end of V_L as described by Richardson et al. 1995. To prepare for the construction of a recombinant AAV vector, an XbaI restriction site is incorporated into the forward and reverse primers for reamplifying the recombinant scFv gene so as to facilitate its insertion into the AAV plasmid vector pSSV discussed immediately below.

Construction of recombinant adeno associated viral vectors for regional expression of scFv $\alpha\beta$ genes in the brain

The assembled ScFv $\alpha\beta$ genes are ligated into an AAV plasmid pSSV9 (psub201) under the control of the human β APP promoter (hu β APPP). Plasmid pSSV9 is a modified full-length AAV type 2 genomic clone. Alternatively, other suitable promoters, such as Thy-1, synapsin I, prion, etc. can be used as discussed previously, although hu β APPP is preferred.

As shown in Figure 5, all of the AAV coding sequences are excised, leaving only the viral inverted terminal repeats (ITR) by cleavage of the two flanking XbaI sites. These ITRs contain the recognition signals necessary for replication and packaging into an AAV vector. The AAV coding sequences are replaced with the hu β APPP_{H/K} $\alpha\beta$ coding sequences. The new coding sequences of AAV/hu β APPP_{H/K} $\alpha\beta$ are followed by an SV40 early region polyadenylation signal.

Preparation of packaged hu β APPP_{H/K} $\alpha\beta$ AAV vectors

hu β APPP_{H/K} $\alpha\beta$ AAV vectors are packaged by co-complementation as described by Samulski et al., 1989, using an adenovirus-transformed human embryonic kidney cell line, 293 (ATCC CRL-1573). The cells are cultured in Eagle's MEM supplemented with glutamine and Earle's salts and 10% heat-inactivated fetal calf serum at 37°C in a humidified incubator with 5% CO₂. Adenovirus type 5 (Ad5) stocks are raised by 1 h infection of subconfluent 293 cells with 10 μ l of Ad5 seed culture in 1 ml of serum-free DMEM per 100 mm dish. After 48 to 72 h, when strong cytopathic effects are observed, the cells are collected and pelleted by centrifugation. The cells are lysed by freeze/thaw cycles to

release the intracellular virus and debris is removed by low-speed centrifugation. Virus containing supernatants are aliquoted and stored -70°C. Co-complementation is accomplished as follows: Semi-confluent 293 cells plated at 0.5 x 10⁶ cells per 100 mm plate are infected with Ad5 at a multiplicity of infection of 10. After 1 h, the cells are co-transfected with 20 µg of huβAPPP_{H/K}αAβ AAV plasmids and 10 µg of pAd8 which contains the AAV 2 genes encoding the AAV replication and encapsidation functions, but flanked by terminal repeats that are derived from adenovirus 2, rather than AAV (Samulski et al., 1987, 1989). Transfection is performed using a standard calcium phosphate precipitation method (Wigler et al., 1979) with the addition of chloroquine diphosphate to enhance the transfection efficiency (Luthman et al., 1983). After overnight incubation, the transfection solution is replaced with fresh medium containing 15% fetal calf serum. Three days after infection the cells are harvested, pelleted by centrifugation at 1000 x g for 10 min, and then subjected to freeze thaw cycles to release the cell-associated virions. Contaminating Ad5 is inactivated by heating lysates to 56°C. The lysates are then clarified by centrifugation and treated with 25 units/ml of RNASE-free DNASE at 37°C for 30 min to remove any remaining plasmid DNA.

Transduction of human neurons to test expression of AAV/huβAPPP_{H/K}αAβ vectors

The utility of AAV as a potential vector has been established unequivocally by Du et al., 1996, in human NT neurons (Pleasure et al., 1993). The precursor of these neurons is a subline of human teratocarcinoma cells, NT2, that commits to terminal differentiation into neurons on exposure to retinoic acid (Lee et al., 1994; Pleasure et al., 1993). Four weeks of retinoic acid treatment accompanied by selective replatings can yield nearly pure > 95 % populations of neurons. These mature neurons remain viable in culture for many weeks. In addition to the distinct morphological

appearance and expression of many neuronal markers, human NT neurons have similar patterns of amyloid precursor proteins as native CNS neurons and produce A β peptides (Wertkin et al., 1993).

- 5 Undifferentiated precursor NT2 cells are obtained from Stratagene, La Jolla, CA, and cultured in Opti-MEM (GIBCO BRL, Gaithersburg, MD) containing 5% heat-inactivated fetal bovine serum (FBS) and 100 units/ml penicillin and 100 μ g/ml streptomycin (PS) at 37°C in 5% CO₂. 2 x 10⁶ NT2 cells
10 per T75 flask are treated with 10 μ M retinoic acid for four weeks and then replated at low density into six T75 flasks. The top layers containing differentiated cells are mechanically dislodged and replated at 1 x 10⁶ cells per well in 24-well plates. Wells and glass cover-slips are coated
15 with 0.01% poly-D-lysine followed by 1:20 MATRIGEL (Collaborative Research, Bedford, MA). Cells are cultured in DMEM high glucose/L-glutamine containing 10% FBS, PS and mitotic inhibitors for three weeks. The enriched neurons are maintained in DMEM/10% FBS/PS at 37°C, 5% CO₂.
- 20 NT neurons (about 10⁵ per well) are transduced with AAV/hu β APP_{H/K}A β vectors by removing the growth medium, washing once with serum-free medium and adding vector stock diluted in serum-free DMEM. After incubating for 90 min at 37°C, 1 ml of DMEM containing 10% FBS is added to each well.
25 The cultures receive a change of medium after two days, and twice weekly thereafter.

Cell viability assay

- 30 The viability of cells is evaluated on the basis of the mitochondrial function of control and hu β APP_{H/K}A β AAV vector transduced cells. The levels of mitochondrial dehydrogenase activity were compared using 3-(4,5-Dimethylthiazol-3-yl)-2,5-diphenyl tetrazolium bromide as the substrate. Its cleavage to a purple formazan product
35 by dehydrogenase is spectrophotometrically quantified at 570 nm.

Detection and determination of binding affinity constants

To verify that the secreted recombinant antibodies (antisenilins) retain the binding properties of the original hybridoma secreted antibodies, ELISA assays are performed, as described earlier in this example, with the culture medium of NT2 transduced cells.

Bioassays to test inhibition of A β functions

The secreted antibodies are isolated from the culture medium in which the transduced NT2 cells are incubated. The purified antibodies and the culture-medium itself are tested as inhibitors of A β aggregation or A β induced cytotoxicity as described above in the section on *in vitro* bioassays.

Generation of transgenic mice expressing the single-chain antisenilin antibodies in the brain

The ScFv antisenilin gene is inserted into a hamster prion protein (PrP) cosmid vector in which the PrP open reading frame (ORF) is replaced with the antisenilin gene ORF. The transgenes are used to generate transgenic mice by microinjection into fertilized 1-cell eggs of C57B6SJL mice according to any one of the widely used methods, such as Brinster et al. (1981), Harbers et al. (1981), Wagner et al. (1981), Gordon et al. (1976), Stewart et al. (1982), Palmiter et al. (1983), and U.S. Patent No. 4,870,009. The resulting progeny (TGScFvA) are tested by genotyping using standard PCR amplification procedures.

Animal models to establish the therapeutic potential of the $\alpha\beta$ antibodies as antisenilins

Animal models are required to test for the expression of anti-A β antibodies *in vivo* and to determine whether they demonstrate a potential for slowing down the accumulation of amyloid plaques and prevent the development of AD-like pathology in the brain. Although AD is a uniquely human disease, a number of transgenic mice that overexpress human β APP show promise.

Effects of chronic A β depletion on plaque burden and related AD pathology transgenic mice

The antisenilin function of the recombinant A β end-specific antibodies are tested in a transgenic animal mouse model that overexpresses the 695-amino acid isoform of Alzheimer β APP containing a Lys670 to Asn, Met671 to Leu mutation (Hsiao, K., 1996, U.S. Patent Application no. 08/664,872). The correlative appearance of behavioral, biochemical, and pathological abnormalities reminiscent of Alzheimer's Disease in these transgenic mice (TG2576) provides the opportunity to explore the usefulness of agents to slow down or prevent the A β -induced pathophysiology of the disease.

Female transgenic mice (TGScFvA) homozygous for the antisenilin gene are crossed with breeding TG2576 males. The offspring that express both the antisenilin gene and the variant APP gene are compared with respect to behavioral, biochemical, and pathological abnormalities with TG2576 mice.

Having now fully described this invention, it will be appreciated by those skilled in the art that the same can be performed within a wide range of equivalent parameters, concentrations, and conditions without departing from the spirit and scope of the invention and without undue experimentation.

While this invention has been described in connection with specific embodiments thereof, it will be understood that it is capable of further modifications. This application is intended to cover any variations, uses, or adaptations of the inventions following, in general, the principles of the invention and including such departures from the present disclosure as come within known or customary practice within the art to which the invention pertains and as may be applied to the essential features hereinbefore set forth as follows in the scope of the appended claims.

All references cited herein, including journal articles or abstracts, published or unpublished U.S. or foreign patent applications, issued U.S. or foreign patents,

or any other references, are entirely incorporated by reference herein, including all data, tables, figures, and text presented in the cited references. Additionally, the entire contents of the references cited within the references cited herein are also entirely incorporated by reference.

Reference to known method steps, conventional methods steps, known methods or conventional methods is not in any way an admission that any aspect, description or embodiment of the present invention is disclosed, taught or suggested in the relevant art.

The foregoing description of the specific embodiments will so fully reveal the general nature of the invention that others can, by applying knowledge within the skill of the art (including the contents of the references cited herein), readily modify and/or adapt for various applications such specific embodiments, without undue experimentation, without departing from the general concept of the present invention. Therefore, such adaptations and modifications are intended to be within the meaning and range of equivalents of the disclosed embodiments, based on the teaching and guidance presented herein. It is to be understood that the phraseology or terminology herein is for the purpose of description and not of limitation, such that the terminology or phraseology of the present specification is to be interpreted by the skilled artisan in light of the teachings and guidance presented herein, in combination with the knowledge of one of ordinary skill in the art.

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(ii) TITLE OF INVENTION: DNA ENCODING RECOMBINANT ANTIBODY MOLECULES
END-SPECIFIC FOR AMYLOID-BETA PEPTIDES, PHARMACEUTICAL
COMPOSITIONS THEREOF, AND METHOD OF PREVENTING OR INHIBITING
PROGRESSION OF ALZHEIMER'S DISEASE

(iii) NUMBER OF SEQUENCES: 5

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(v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30

(vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:
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(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 59 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Asp Ala Glu Phe Arg Cys
1 5

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

Asp Ala Glu Phe Arg His Asp Cys
1 5

(2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Cys Leu Met Val Gly Gly Val Val
1 5

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Cys Val Gly Gly Val Val Ile Ala
1 5

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
- Asp Ala Glu Phe Arg His
1 5
- (2) INFORMATION FOR SEQ ID NO:7:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 13 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
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1 5 10
- (2) INFORMATION FOR SEQ ID NO:8:
- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
- Glu Phe Arg His
1

WHAT IS CLAIMED IS:

1. A method for preventing or inhibiting progression of Alzheimer's Disease, comprising the step of administering a composition comprising a recombinant DNA molecule, containing a gene encoding a recombinant antibody molecule end-specific for the N-terminus or the C-terminus of an amyloid- β peptide, operably-linked to a promoter which is expressed in the central nervous system, in association with a means for gene delivery, to a patient in need thereof to prevent the accumulation of amyloid- β peptides and the aggregation of peptides which form amyloid deposits in the brain.

2. The method according to claim 1, wherein the composition is administered by injection intravenously, intra-arterially, intracranially, or intracephalically.

3. The method according to claim 1, wherein the amyloid- β peptide is selected from the group consisting of amyloid β -peptides having the amino acid sequence of residues 5-44 of SEQ ID NO:1, residues 5-46 of SEQ ID NO:1, residues 5-47 of SEQ ID NO:1, and mixtures thereof.

4. The method according to claim 1, wherein the recombinant antibody molecule is end-specific for the N-terminus of the amyloid- β peptide.

5. The method according to claim 1, wherein the recombinant antibody molecule is end-specific for the C-terminus of the amyloid- β peptide.

6. The method according to claim 1, wherein the promoter operably-linked to the gene encoding a recombinant antibody molecule is a β APP promoter.

7. The method according to claim 1, wherein the means for gene delivery in association with the recombinant DNA molecule comprises a viral vector.

8. The method according to claim 7, wherein the viral vector is adeno-associated vector (AAV).

9. The method according to claim 7, wherein the means for gene delivery further comprises cationic lipids or cationic liposomes.

10. The method according to claim 1, wherein the means for gene delivery in association with the recombinant DNA molecule comprises cationic lipids or cationic liposomes.

11. The method according to claim 1, wherein the means for gene delivery in association with the recombinant DNA molecule comprises a ligand capable of binding to a cell surface receptor.

12. The method according to claim 11, wherein the ligand is biotin.

13. The method according to claim 1, wherein the recombinant antibody molecule is a single chain variable region fragment.

14. Monoclonal antibody end-specific for the N-terminus of an amyloid β -peptide.

15. A recombinant DNA molecule, comprising a gene encoding a recombinant antibody molecule end-specific for the N-terminus or the C-terminus of an amyloid- β peptide and a promoter operably linked to said gene, wherein said promoter is capable of expressing said recombinant antibody molecules in brain cells.

16. The recombinant DNA molecule according to claim 15, wherein said promoter is a β APP promoter.

17. A vector comprising the recombinant DNA molecule of claim 15.

18. A host cell transformed with the vector of claim 17.

19. A pharmaceutical composition for preventing or inhibiting progression of Alzheimer's Disease, comprising the recombinant DNA molecule of claim 15 in association with a means for gene delivery, and a pharmaceutically acceptable excipient.

20. The pharmaceutical composition according to claim 19, wherein the means for gene delivery is selected from the group consisting of viral vectors, cationic lipids, cationic liposomes, ligands capable of binding to a cell surface receptor, and combinations thereof.

21. The pharmaceutical composition according to claim 19, wherein said gene encodes a recombinant antibody

molecule end-specific for the N-terminus of an amyloid- β peptide.

22. The pharmaceutical composition according to claim 19, wherein said gene encodes a recombinant antibody molecule end-specific for the C-terminus of an amyloid- β peptide.

ABSTRACT

DNA encoding a recombinant antibody molecule end-specific for an amyloid-beta peptide, pharmaceutical compositions thereof, and a method for preventing or inhibiting progression of Alzheimer's Disease by introducing such a DNA molecule into brain cells to express the recombinant antibody molecule and prevent the accumulation of amyloid-beta peptides in the cerebrospinal fluid.

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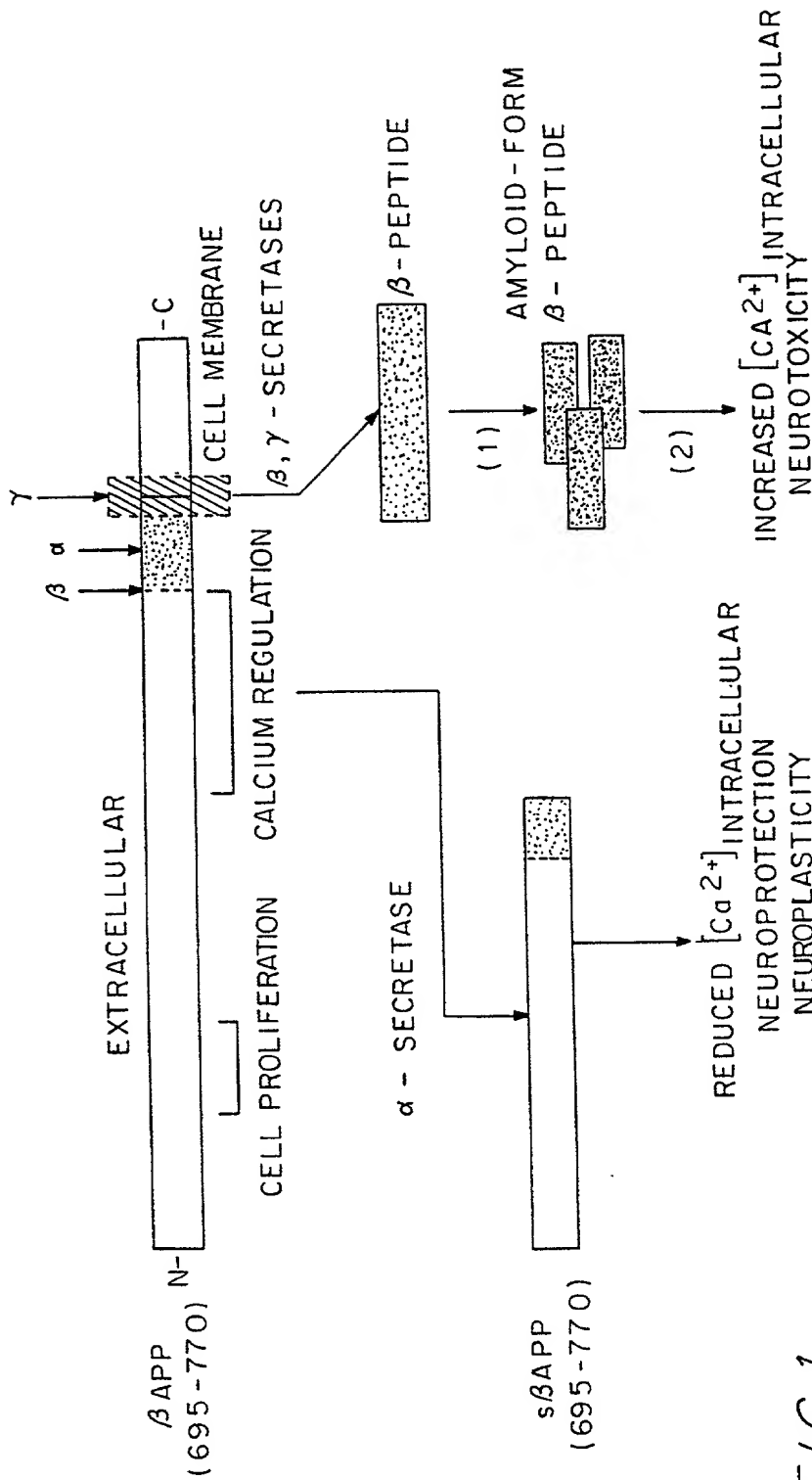
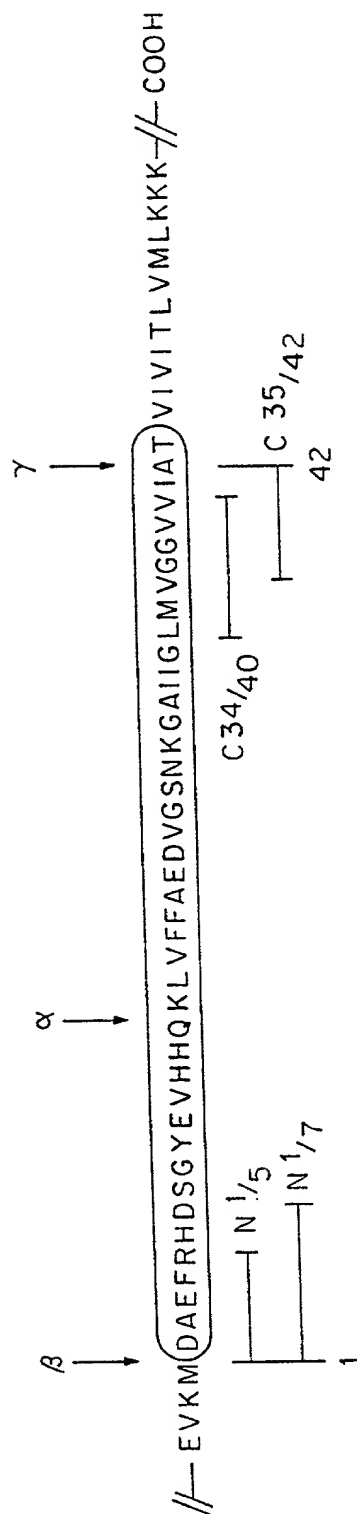


FIG. 1

FIG. 2



ANTIBODY

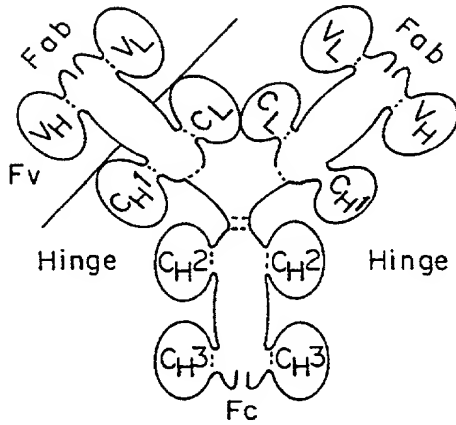


FIG. 3A

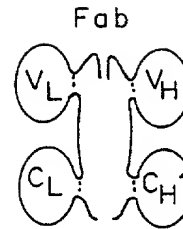


FIG. 3B

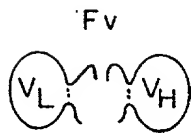
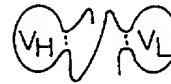


FIG. 3C

SINGLE CHAIN Fv (ScFv)



H₂N - V_H - LINKER - V_L - COOH

FIG. 3D

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FIG. 4

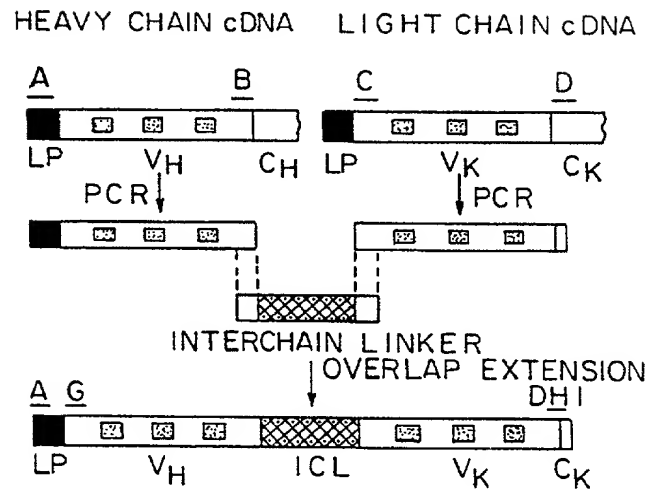
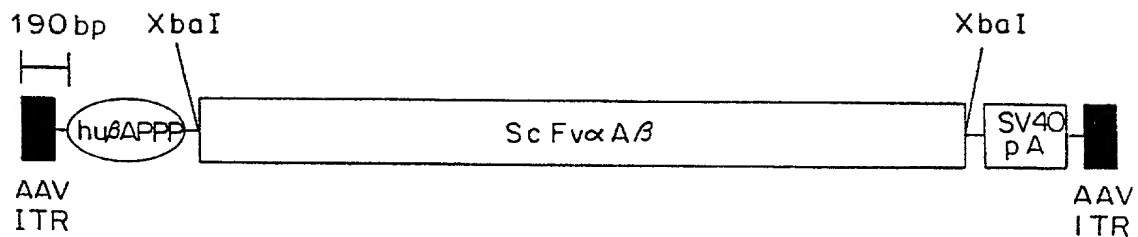


FIG. 5



Combined Declaration for Patent Application and Power of Attorney

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name; and that I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

RECOMBINANT ANTIBODIES SPECIFIC FOR BETA-AMYLOID ENDS, DNA ENCODING AND METHODS OF USE THEREOF the specification of which (check one)

☐ is attached hereto;

☐ was filed in the United States under 35 U.S.C. §111 on _____, as
USSN _____; or

☒ was/will be filed in the U.S. under 35 U.S.C. §371 by entry into the U.S. national stage of
an international (PCT) application, PCT/US98/06900; filed 09 April 1998,

entry requested on _____; national stage application received

USSN _____; §371/§102(e) date _____ (*if known),

and was amended on _____ (if applicable).

(include dates of amendments under PCT Art. 19 and 34 if PCT)

I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above; and I acknowledge the duty to disclose to the Patent and Trademark Office (PTO) all information known by me to be material to patentability as defined in 37 C.F.R. § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. §§ 119, 365 of any prior foreign application(s) for patent or inventor's certificate, or prior PCT application(s) designating a country other than the U.S., listed below with the "Yes" box checked and have also identified below any such application having a filing date before that of the application on which priority is claimed:

(Number)	(Country)	(Day Month Year Filed)	<input type="checkbox"/>	<input type="checkbox"/>
			YES	NO
(Number)	(Country)	(Day Month Year Filed)	<input type="checkbox"/>	<input type="checkbox"/>
			YES	NO

I hereby claim the benefit under 35 U.S.C. § 120 of any prior U.S. non-provisional Application(s) or prior PCT Application(s) designating the U.S. listed below, or under § 119(e) of any prior U.S. provisional applications listed below, and, insofar as the subject matter of each of the claims of this application is not disclosed in such U.S. or PCT application in the manner provided by the first paragraph of 35 U.S.C. §112, I acknowledge the duty to disclose to the PTO all information as defined in 37 C.F.R. §1.56(a) which occurred between the filing date of the prior application and the national filing date of this application:

<u>60/041,850</u>	<u>09 April 1997</u>	<u>abandoned</u>
(Application Serial No.)	(Day Month Year Filed)	(Status: patented, pending, abandoned)
_____	_____	_____
(Application Serial No.)	(Day Month Year Filed)	(Status: patented, pending, abandoned)
_____	_____	_____
(Application Serial No.)	(Day Month Year Filed)	(Status: patented, pending, abandoned)

I hereby appoint the following attorneys, with full power of substitution, association, and revocation, to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith.

SHERIDAN NEIMARK, REG. NO. 20,520 - ROGER L. BROWDY, REG. NO. 25,618 - ANNE M. KORNBAU, REG. NO. 25,884
NORMAN J. LATKER, REG. NO. 19,963 - IVER P. COOPER, REG. NO. 28,005 - ALLEN C. YUM, REG. NO. 37,921*
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DIRECT ALL TELEPHONE CALLS TO:
BROWDY AND NEIMARK
(202) 628-5197

The undersigned hereby authorizes the U.S. Attorneys or Agents named herein to accept and follow instructions from MINDSET LTD. as to any action to be taken in the U.S. Patent and Trademark Office regarding this application without direct communication between the U.S. Attorney or Agent and the undersigned. In the event of a change of the persons from whom instructions may be taken, the U.S. Attorneys or Agents named herein will be so notified by the undersigned.

Title: DNA ENCODING RECOMBINANT ANTIBODY MOLECULES END-SPECIFIC FOR AMYLOID-BETA...

U.S. Application filed _____, Serial No. _____

PCT Application filed 09 April 1998, Serial No. PCT/US98/06900

I hereby further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C. §1001 and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

FULL NAME OF FIRST INVENTOR Daniel G. CHAIN		INVENTOR'S SIGNATURE <i>Daniel Chain</i>	DATE 10-06-99
RESIDENCE Old Katamon, Israel <i>TLX</i>		CITIZENSHIP Great Britain	
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FULL NAME OF SECOND JOINT INVENTOR		INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP	
POST OFFICE ADDRESS			
FULL NAME OF THIRD JOINT INVENTOR		INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP	
POST OFFICE ADDRESS			
FULL NAME OF FOURTH JOINT INVENTOR		INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP	
POST OFFICE ADDRESS			
FULL NAME OF FIFTH JOINT INVENTOR		INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP	
POST OFFICE ADDRESS			
FULL NAME OF SIXTH JOINT INVENTOR		INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP	
POST OFFICE ADDRESS			
FULL NAME OF SEVENTH JOINT INVENTOR		INVENTOR'S SIGNATURE	DATE
RESIDENCE		CITIZENSHIP	
POST OFFICE ADDRESS			

ALL INVENTORS MUST REVIEW APPLICATION AND DECLARATION BEFORE SIGNING. ALL ALTERATIONS MUST BE INITIALED AND DATED BY ALL INVENTORS PRIOR TO EXECUTION. NO ALTERATIONS CAN BE MADE AFTER THE DECLARATION IS SIGNED. ALL PAGES OF DECLARATION MUST BE SEEN BY ALL INVENTORS.

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